# Enterotoxinproducing Escherichia coli 0169:H41, United States

Mark E. Beatty,\* Cheryl A. Bopp,\* Joy G. Wells,\*
Kathy D. Greene,\* Nancy D. Puhr,\*
and Eric D. Mintz\*

From 1996 to 2003, 16 outbreaks of enterotoxigenic *Escherichia coli* (ETEC) infections in the United States and on cruise ships were confirmed. *E. coli* serotype O169:H41 was identified in 10 outbreaks and was the only serotype in 6. This serotype was identified in 1 of 21 confirmed ETEC outbreaks before 1996.

Enterotoxigenic *Escherichia coli* (ETEC) is an important cause of diarrhea in the developing world and among travelers and is increasingly recognized as a cause of outbreaks in the United States (1). Dalton et al. reviewed confirmed ETEC outbreaks that occurred both in the United States and among passengers on cruise ships that docked in U.S. ports from 1975 through 1995 (2). Twenty-one such outbreaks caused by 17 different ETEC serotypes occurred during this period; 7 (33%) occurred among cruise ship passengers (2). Because laboratory tests for the identification of ETEC are not widely available, outbreaks caused by ETEC may escape recognition, and healthcare workers may miss opportunities for treatment and prevention. To improve recognition of ETEC outbreaks, Dalton proposed that specimens from outbreaks of gastroenteritis that meet certain criteria be referred for ETEC testing at a public health reference laboratory. These outbreaks include those for which routine stool cultures have not yielded an etiologic agent and those which are characterized by an incubation period of 24 to 48 hours, a duration of illness >60 hours, and a diarrhea-to-vomiting prevalence ratio of  $\geq 2.5$  (2).

## The Study

For the 8-year period 1996 through 2003, we reviewed all suspected ETEC outbreaks solely or jointly investigated by the Foodborne and Diarrheal Diseases Branch at the Centers for Disease Control and Prevention (CDC). In accordance with Dalton et al., we defined a confirmed

ETEC outbreak as one in which ETEC isolates of the same serotype were isolated from  $\geq 3$  ill persons and no other viral or bacterial pathogens were identified, or one in which ETEC isolates of the same serotype were isolated from  $\geq 10$  ill persons and no more than one other bacterial or viral pathogen was identified in a single stool specimen. Stool specimens collected during these investigations were routinely cultured for *Salmonella*, *Shigella*, *Campylobacter*, *E. coli* O157:H7, *Yersinia*, and *Vibrio* spp. In many instances, these specimens were also tested for noroviruses.

To identify ETEC, patient specimens were plated to MacConkey agar, and individual colonies or sweeps of confluent growth were tested by polymerase chain reaction (PCR) for heat-labile (LT) and heat-stable (ST) enterotoxin genes (3). Using standard methods, we serotyped LT- or ST-positive isolates for O and H antigens. ETEC isolates were tested by the disk-diffusion method for susceptibility to ampicillin, amoxicillin/clavulanic acid, ceftriaxone, chloramphenicol, ciprofloxacin, gentamicin, kanamycin, nalidixic acid, streptomycin, sulfisoxazole, tetracycline, and trimethoprim-sulfamethoxazole (4). A PCR–restriction fragment length polymorphism test was used to identify the H41 gene in selected nonmotile *E. coli* O169 isolates (5).

## **Conclusions**

During the 8-year study period, CDC received isolates from 59 outbreaks for ETEC testing. Sixteen met the criteria for our definition of a confirmed ETEC outbreak; three occurred on international cruise ships that docked in U.S. ports and 12 occurred in the United States (Table). We identified ETEC in specimens from six other outbreaks that did not meet these criteria, either because ETEC isolates of the same serotype were isolated from only two persons (n = 2) or because we isolated additional bacterial pathogens from the specimens (n = 4).

The 16 ETEC outbreaks had a median of 41 ill persons per outbreak (range 5-916), for a total of 2,865 patients. From 81% to 100% of ill persons in each outbreak reported diarrhea; less frequently reported symptoms included abdominal cramps (66%-90%), fever (0%-73%), nausea (44%-70%), and vomiting (0%-33%). In the 15 outbreaks in which diarrhea and vomiting were reported, the median "diarrhea-to-vomiting prevalence ratio" (the percentage of patients who reported diarrhea divided by the percentage of patients who reported vomiting) was 7.7 (range 2.9-undefined: the upper limit of the range is undefined because in two outbreaks all ill persons interviewed denied vomiting). In nine outbreaks with sufficient data, incubation periods among individual patients were 5–158 hours. The median incubation period was 24-48 hours for 10 of the 12 outbreaks for which it could be calculated. The

<sup>\*</sup>Centers for Disease Control and Prevention, Atlanta, Georgia, USA

Table. Characteristics of enterotoxigenic Escherichia coli (ETEC) outbreaks, a United States, 1996–2003

			G 44			%		Median	Median illness			
No.	Mo/y	Location	Setting (reference)	Presumed source	No. ill	Diarrhea (bloody)	% Vomiting	incubation, h (range)	duration, days (range)	No isolates	Serotype/toxin type <sup>b</sup>	Antimicrobial resistance <sup>c</sup>
1	3/96	Caribbean	Cruise ship	Drinking water	652	98 (9)	19	_	_	6	O169:H41/ST	Тс
										1	O6:H16/ LT,ST	Sensitive
										1	O27:H7/ST	St, Su, Tc
										1	O34:H10/ST	Tc
2	4/97	Caribbean	Cruise ship	Drinking water,	429	100 (6)	14	_	>3	8	O169:H41/ST	Tc
			(5)	ice						3	O148:H28/ LT,ST	Тс
										1	O78:H12/ST	Ch, St, Su, Tc, TmS
										1	O27:H7/ST	St, Su, Tc
3	4/97	California	Restaurant	Beans, enchilada, tacos, rice, tortilla chips	41	95	17	24 (5–85)	2 (0.3–4)	11	O27:H7/ST	Sensitive
4	6/97	Massachusetts	Boxed lunch	Tomato, mozzarella salad	33	97	33	48 (24–96)	3.7 (1–7)	5	O25:NM/ST	Sensitive
5	7/97	Minnesota	Catered party	Fresh vegetables	15	100	13	37 (8-158)	3.2 (0.4–6)	4	O169:H41/ST	Tc
6	4/98	Mexico-	Cruise ship	_	397	96	33	-	-	3	O6:H16/ LT,ST	Sensitive
		Hawaii								1	O6:H16/ LT,ST	St, Tc
										2	O169:H41/ST	Tc
										1	O148:H28/ LT,ST	St, Su, Tc
										1	O148:H28/ LT,ST	Тс
										1	O27:H7/ST	St, Su, Tc
7	6/98	Illinois	Catered parties	Potato, macaroni, egg salads	916	100	8	50 (40–76)	5 (2–9)	11	O6:H16/ LT,ST	Sensitive
8	8/98	Minnesota	Restaurant (6)	Parsley	66	100 (0)	6	25	8	7	O6:H16/ LT,ST	Ap, St, Su, Tc
										1	O159:H4/LT	Ap, St, Su, Tc, TmS
										1	O27:H7/ST	St, Su, Tc
9	9/98	Minnesota	Restaurant	_	5	100(0)	0	48	6	3	O169:H41/ST	St, Su, Tc
10	5/00	Washington	Cruise ship	Basil	100	100	5	40 (27–67)	10 (0.5–21)	3	O169:H41/ST	Тс
11	6/00	New York	Banquet	-	40	97 (0)	3	48 (24–96)	3 (1–8)	5	O169:H41/ST	Tc
12	7/00	Utah	Wedding rehearsal	_	45	100	29	33 (18–59)	2.3 (1–3)	5	O27:H7/ST	St, Su, Tc
13	7/01	Wisconsin	Catered party	Quesadillas, fajitas, nacho chips, beans	21	100	-	38 (9–70)	6 (1–8)	3	O169:H41/ST	Тс
14	8/01	Illinois	Catered	_	24	100	0	-	-	3	O169:H41/ST	Ap, Tc
			party							1	O169:H41/ST	Tc
										2	O6:H16/LT,ST	Sensitive
										1	O25:NM/ST	Ap
15	10/02	Oregon	Catered party	Garlic chicken lasagna	40	98 (13)	15	72 (24–144)	5	3	O27:H7/ST	St, Su,Tc
16	8/03	Tennessee	Catered party	Catfish, coleslaw	41	81 (0)	5	2	2.5	12	0169:H49	Tc

 $<sup>^{</sup>a}$ An outbreak is defined as  $\geq 3$  ill persons infected with the same ETEC serotype and no other viral or bacterial pathogens, or  $\geq 10$  ill persons with the same serotype and no more than one other bacterial or viral pathogen identified.

duration of illness, reported in 11 outbreaks, was 0.3-21 days. The median duration of illness was  $\geq$ 60 hours in 11 of the 13 outbreaks for which it could be calculated.

A vehicle was implicated in 11 (69%) outbreaks. Unbottled ship's water or beverages containing ice prepared on board the ship were implicated in 2 of the 3 outbreaks on cruise ships that had docked in foreign ports.

The source of the third international cruise ship outbreak was not determined. Although no problems with chlorination of bunkered water were documented in the two outbreaks, this problem has been seen in previous waterborne ETEC outbreaks aboard cruise ships (6). Basil served on board ship was implicated as the source of ETEC in the remaining cruise ship outbreak (on a ship that docked only

<sup>&</sup>lt;sup>b</sup>NM, nonmotile; LT, heat-labile toxin; ST, heat-stable toxin.

<sup>&</sup>lt;sup>c</sup>Ap, ampicillin; Ch, chloramphenicol; St, streptomycin; Su, sulfisoxazole; Tc, tetracycline; TmS, trimethoprim-sulfamethoxazole

in U.S. ports). One other outbreak was attributed to a fresh herb (parsley) served raw (7). Salads made with raw vegetables were implicated in four other domestic outbreaks.

If we define a strain as each ETEC serotype identified during an outbreak that has a unique antimicrobial resistance pattern, we identified a total of 30 strains representing eight different serotypes in specimens from the 16 outbreaks (Table). In five outbreaks, we isolated more than one ETEC serotype. Heat-stable toxin (ST)-producing E. coli O169:H41 was the most commonly identified serotype. This serotype was identified as the only pathogen in specimens from six outbreaks in the United States and was identified along with other ETEC serotypes in four additional outbreaks. Three of these four outbreaks occurred on international cruise ships. In 21 previously reported ETEC outbreaks, E. coli O169:H41 had been isolated only once, from an outbreak that occurred among international cruise ship passengers in 1995, in which another ETEC serotype predominated (2). By all of the basic epidemiologic and clinical characteristics that we analyzed, outbreaks in which E. coli O169:H41 was identified alone, or in combination with other serotypes, did not appear to differ from outbreaks in which this emerging strain was not identified.

Resistance to antimicrobial agents remained common among ETEC isolates (Table). Twenty-four (80%) of the 30 strains were resistant to tetracycline, 11 (38%) were resistant to sulfisoxazole, 4 (13%) were resistant to ampicillin, and 2 (7%) were resistant to trimethoprim-sulfamethoxazole. All strains of O169:H41 were resistant to tetracycline, and two were also resistant to at least one additional antimicrobial drug. Only two outbreaks were caused exclusively by pan-sensitive ETEC strains (7%).

A comparison of ETEC outbreaks reported to CDC from 1996 through 2003 with those from previous years shows that outbreaks on cruise ships and in the United States continue to occur and that antimicrobial resistance among ETEC isolates remains common. Raw vegetables and herbs have been increasingly implicated as the vehicles for ETEC outbreaks in recent years. This finding is in keeping with an increase in produce-associated outbreaks among other foodborne bacterial pathogens (8). Finally, a new ETEC serotype, ST-producing O169:H41, has become predominant.

The first report of O169:H41 was in association with a foodborne outbreak in 1991 in Japan, where this serotype continues to be isolated (9–11). Hamada reported four outbreaks that occurred from June 1997 to August 1998; the largest of these outbreaks had a 57% attack rate and resulted in approximately 2,800 cases. All four outbreaks occurred at either restaurants or catered events (11). Contaminated wakame seaweed was implicated in one of the outbreaks and considered the likely cause in another

(11). Nishakawa et al. report that O169:H41 has become the most prevalent ETEC serotype in Japan (12).

In 1995, CDC detected this serotype for the first time during an outbreak on a Caribbean cruise ship. It was identified during two additional Caribbean cruise ship outbreaks before causing a domestic outbreak in Minnesota in 1997. In addition to being identified in 10 of the 16 ETEC outbreaks that met our criteria, O169:H41 was also a predominant serotype in 4 of the 6 ETEC outbreaks that did not meet our criteria for a confirmed outbreak, either because the serotype was isolated from two persons only or because an additional pathogen was also isolated in the outbreak.

The emergence and eventual predominance of O169:H41 in the United States and Japan may have important implications for ETEC vaccine producers. Nishikawa et al. characterized strains of O169:H41 from Japan and reported that they are not clonal and that they possess a novel colony-forming factor (12).

From 1996 through 1999, laboratory-confirmed ETEC outbreaks represented 0.2% of all foodborne outbreaks reported to CDC (13). This number is likely to be an underestimate because special diagnostic tests are required to confirm ETEC. Seventy-one percent of outbreaks of foodborne illness reported to CDC during this period were of unknown cause. In a previous study, Hall et al. demonstrated that the epidemiologic and clinical syndrome in 1.1% of outbreaks of unknown cause reported to CDC from 1982 through 1989 was compatible with infections caused by ETEC or STEC (14). ETEC is also responsible for some episodes of sporadic diarrheal disease. When researchers systematically looked for it, they isolated ETEC from 1.4% of stool samples from patients visiting urban and rural health maintenance organization clinics in Minnesota for diarrhea (15). Our data suggest that the clinical criteria proposed by Dalton et al. for suspecting ETEC as a cause of an outbreak of unknown cause (median incubation 24-48 hours, mean or median duration >60 hours, and diarrhea-tovomiting prevalence ratio  $\geq 2.5$ ) remain valid.

The epidemiology of ETEC outbreaks in the United States is changing, but the incidence of these outbreaks does not appear to be decreasing. Researchers cannot use routine stool cultures to detect ETEC, and delays in stool sample collection for  $\geq 7$  days greatly reduces yield (16,17). For outbreaks that meet the clinical profile and for which routine stool diagnostic tests have not yielded an enteric pathogen, physicians and public health authorities should send *E. coli* isolates to reference laboratories, such as CDC, for ETEC testing.

#### **Acknowledgments**

We thank the staff of the local and state health departments and the Vessel Sanitation Program at the Centers for Disease Control and Prevention, who participated in the investigations summarized in this manuscript.

Dr. Beatty, a pediatrician, is a preventive medicine resident at the Centers for Disease Control and Prevention. His research interests include enteric pathogens.

### References

- Cohen MB, Giannella RA. Enterotoxigenic *Escherichia coli*. In: Blaser MJ, Smith PD, Ravdin JI, Greenberg HB, Guerrant RL, editors. Infections of the gastrointestinal tract. 2nd ed. Philadelphia: Lippincott Williams & Wilkins; 2002. p. 579–94.
- Dalton CB, Mintz ED, Wells JG, Bopp CA, Tauxe RV. Outbreaks of enterotoxigenic *Escherichia coli* infection in American adults: a clinical and epidemiologic profile. Epidemiol Infect 1999;123:9–16.
- Olsvik O, Strockbine NA. PCR detection of heat-stable, heat-labile, and Shiga-like toxin genes in *Escherichia coli*. In: Persing CH, Smith TF, Tenover FC, White TJ, editors. Diagnostic molecular microbiology. Washington: American Society for Microbiology; 1993. p. 271–6
- National Committee for Clinical Laboratory Standards. Performance standards for antimicrobial disk susceptibility tests. Approved Standard. 5th ed. NCCLS doc. M2-A5 (ISBN 1-56238-208-X). Villanova (PA): The Committee; 1993.
- Fields PI, Blom K, Hughes HJ, Helsel LO, Feng P, Swaminathan B. Molecular characterization of the gene encoding H antigen in *Escherichia coli* and development of a PCR-restriction fragment length polymorphism test for identification of *E. coli* O157:H7 and O157:NM. J Clin Microbiol 1997;35:1066–70.
- Daniels NA, Niemann J, Karpati A, Parashar UD, Greene KD, Wells JG, et al. Traveler's diarrhea at sea: three outbreaks of waterborne enterotoxigenic *Escherichia coli* (ETEC) on cruise ships. J Infect Dis 2000;181:1491–5.
- Naimi TS, Wicklund JH, Olsen SJ, Krause G, Wells JG, Bartkus JM, et al. Concurrent outbreaks of *Shigella sonnei* and enterotoxigenic *Escherichia coli* infections associated with parsley: implications for surveillance and outbreak control. J Food Prot 2003:66:535–41.

- Tauxe RV, Kruse H, Hedberg CW, Potter M, Madden J, Wachmuth K. Microbial hazards and emerging issues associated with produce. J Food Prot 1997;60:1400–8.
- Ando K, Itaya T, Aoki A, Saito A, Masaki H, Tokumura Y, et al. An outbreak of food poisoning caused by enterotoxigenic *E. coli* O169:H41. Jpn J Food Microbiol 1993;10:77–81.
- 10. Nishikawa Y. Heat-stable enterotoxin producing *E. coli* O169:H41 in Japan. Emerg Infect Dis 1995;1:61.
- Hamada K, Tsuji H, Shimada K, Aoki Y. Outbreaks of heat stable enterotoxin-producing *E. coli* O169 in Kinki district in Japan: epidemiological analysis by pulsed-field electrophoresis. Jpn J Infect Dis 1999;52:165–7.
- Nishikawa Y, Helander A, Ogasawara J, Moyer NP, Hanaoka M, Hase A, Yasukawa A. Epidemiology and properties of heat-stable enterotoxin producing *Escherichia coli* serotype O169:H41. Epidemiol Infect 1998;121:31–42.
- Centers for Disease Control and Prevention. U.S. foodborne disease outbreaks. 2002 [cited 2003 Jan 21]. Available from: http://www.cdc.gov/ncidod/dbmd/outbreak/us\_outb.htm.
- Hall JA, Goulding JS, Bean NH, Tauxe RV. Epidemiologic profiling: evaluating foodborne outbreaks for which no pathogen was isolated by routine laboratory testing: United States, 1982-9. Epidemiol Infect 2001;127:381-7.
- 15. Gahr P, Sullivan M, Smith K, Besser J, Hedberg C. A case-control study of enterotoxigenic E. coli infections in Minnesota residents. In: Abstracts of the 41st Interscience Conference on Antimicrobial Agents and Chemotherapy; Chicago, Illinois; 2001 Dec 16–19; Abstract 766. Washington: American Society for Microbiology; 2001.
- Gorbach SL, Kean BH, Evans DG, Bessudo D. Travelers' diarrhea and toxigenic *Escherichia coli*. N Engl J Med 1975;292:933–6.
- Ryder RW, Sack DA, Kapikian AZ, McLaughlin JC, Chakraborty J, Mizanur Rahman AS, et al. Enterotoxigenic *Escherichia coli* and Reo-virus-like agent in rural Bangladesh. Lancet 1976;1:659–63.

Address for correspondence: Mark E. Beatty, Centers for Disease Control and Prevention, Mailstop A38, 1600 Clifton Road, Atlanta, Georgia USA, 30333; fax: 404-639-2205; email: zbn5@cdc.gov

EMERGING Full text free online at www.cdc.gov/eid	
INFECTIOUS DISEASES  The print journal is available at no charge to public health professionals	
YES, I would like to receive Emerging Infectious Diseases.	
Please print your name and business address in the box and return by fax to 404-371-5449 or mail to EID Editor CDC/NCID/MS D61 1600 Clifton Road, NE Atlanta, GA 30333	
Moving? Please give us your new address (in the box) and print the number of your old mailing label here	